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Serum level of adiponectin is a surrogate independent biomarker of radiographic disease progression in early rheumatoid arthritis: results from the ESPOIR cohort

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Abstract

Introduction: Adipokines such as adiponectin, leptin, and visfatin/nicotinamide phosphoribosyltransferase (NAMPT) have recently emerged as pro-inflammatory mediators involved in the pathophysiology of rheumatoid arthritis (RA). We aimed to determine whether serum adipokine levels independently predicted early radiographic disease progression in early RA.

Methods: In total, 791 patients were included from the prospective Etude et Suivi des POLyarthrites Indifférenciées Récentes (ESPOIR) cohort who met the American College of Rheumatology-European League Against Rheumatism criteria for RA ($n = 632$) or had undifferentiated arthritis (UA) ($n = 159$). Enzyme-linked immunosorbent assay (ELISA) was used to assess baseline serum levels of adiponectin, leptin, and visfatin/NAMPT. In the RA group, we tested the association of serum adipokine levels and (a) baseline radiographic damage and (b) radiographic disease progression, defined as a change >0 or ≥ 5 in total Sharp-van der Heijde Score (Δ SHS) between inclusion and 1 year (Δ SHS ≥ 1 or rapid radiographic progression: Δ SHS ≥ 5), adjusting for confounders (age, sex, body-mass index, insulin resistance, C-reactive protein level, Disease Activity Score in 28 joints, Health Assessment Questionnaire score, autoantibody status, steroid use, and radiographic evidence of RA damage at inclusion).

Results: Adiponectin level was independently associated with baseline total SHS (adjusted $\beta = 0.12$; $P = 0.006$). It was also associated with Δ SHS ≥ 1 (adjusted odds ratio (aOR) = 1.84 (1.25 to 2.72)) involving erosive as well as narrowing disease progression (aOR = 1.73 (1.17 to 2.55) and 1.93 (1.04 to 3.57), respectively). Serum adiponectin level predicted Δ SHS ≥ 5 (aOR = 2.0 (1.14 to 3.52)). Serum leptin level was independently associated only with Δ SHS >0 (aOR = 1.59 (1.05 to 2.42)). Conversely, serum visfatin/NAMPT level and radiographic disease progression were unrelated. Considering the receiver-operated characteristic curves, the best adiponectin cut-offs were 4.14 μ g/ml for Δ SHS ≥ 1 and 6.04 μ g/ml for Δ SHS ≥ 5 , with a good specificity (58% and 75% for Δ SHS ≥ 1 and Δ SHS ≥ 5 , respectively) and high negative predictive values (75% and 92% for Δ SHS ≥ 1 or Δ SHS ≥ 5 , respectively).

Conclusion: Serum adiponectin level is a simple useful biomarker associated with early radiographic disease progression in early RA, independent of RA-confounding factors and metabolic status.

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Introduction

In rheumatoid arthritis (RA), early diagnosis and early therapeutic intervention improve clinical outcomes and reduce the accrual of joint damage and major irreversible long-term disability. Patients with early RA should be actively treated as soon as the disease is diagnosed [1]. Finding serum biomarkers that can be used to identify patients at high risk of structural disease progression early in the disease is important because joint lesions occur mainly during this period and may be reduced by early treatment [2]. Levels of inflammatory biomarkers (that is, C-reactive protein (CRP) and erythrocyte sedimentation rate (ESR)) and presence of autoantibodies (that is, rheumatoid factor (RF) and anti-cyclic citrullinated peptide [anti-CCP] antibodies) are associated with subsequent RA structural severity [3-5]. However, the predictive value of these markers for early radiographic disease progression remains weak, especially in early disease, which supports the need for more sensitive and specific biomarkers [6].

Adipokines are adipose tissue-specific soluble proteins also produced by synovium and cartilage, as well as mononuclear blood cells; they are quantifiable in biologic fluids [7]. The most studied are adiponectin, leptin, and visfatin (also known as nicotinamide phosphoribosyltransferase (NAMPT)). These soluble mediators have pleiotropic effects and participate in several metabolic, immune, and inflammatory processes [7-9]. Adiponectin is the most abundant adipokine detected in the serum and has unique properties by acting as an antidiabetic and antiatherogenic mediator [10,11]. However, despite the cardioprotective properties of adiponectin and considering the complexity of its effects, a high serum level of adiponectin was found to be associated with mortality in kidney diseases and type 1 diabetes; this increased level is paradoxically considered as a beneficial physiological response [10-14]. In type 1 diabetes, increased leptin levels are positively associated with β -cell function [15]. In nonrheumatic conditions, serum ratio of adiponectin to leptin may be a noninvasive marker of the severity of nonalcoholic fatty liver disease [16].

The important role of adipokines in inflammation provides novel links between adipose tissue, adipokines, and inflammation-related disorders, including RA [11]. Therefore, adipokines could represent potential disease biomarkers and therapeutic targets in clinical practice. Recent findings have demonstrated that these mediators have potent actions on tissue and cells involved in RA, including cartilage, synovium, bone, and various immune cells [17]. Adipokines as serum biomarkers reflecting RA-related radiographic damage were recently reported in two cross-sectional studies of 167 and 197 subjects [18,19]. Moreover, serum adiponectin level as a predictor of radiographic disease progression has been suggested in two longitudinal studies of 152 patients with established RA and 253 with

early RA [20,21]. However, in all these studies, several important confounding factors, such as metabolic disorders, especially insulin resistance, were not taken into account.

Further to assess the role of adipokines as serum biomarkers for early radiographic disease progression in RA, we determined in a large cohort of patients with early arthritis whether serum adipokine levels (a) were increased in early RA as compared with undifferentiated arthritis (UA), and (b) could predict radiographic structural progression in patients with early RA, and especially rapid radiographic progression.

Methods

Patients

The population and methods of the ESPOIR cohort (Etude et Suivi des POLyarthrites Indifférenciées Récentes) were described in detail elsewhere [22]. Diagnosis of RA was defined as fulfilling the American College of Rheumatology-European League Against Rheumatism (ACR-EULAR) 2010 criteria for RA at inclusion [1]. Otherwise and with no other defined diagnosis at inclusion, patients were considered to have UA. Patients with other defined diagnoses were not assessed for serum adipokine level. The Montpellier (France) Ethics Committee approved the study in July 2002, and all patients provided informed consent.

Serum assays

At inclusion, blood samples were taken for investigation of glycemia, C-reactive protein (CRP) level, and erythrocyte sedimentation rate (ESR). Serum samples were collected at enrolment and immediately stored at -80°C in a single biologic resource centre. A central laboratory was used for determining anti-citrullinated cyclic peptide (anti-CCP) antibodies (anti-CCP2; Dia-Sorin, Saluggia (Vercelli), Italy; positive >50 U/ml) and rheumatoid factor (RF) (Menarini France, Rungis Cedex, France; positive >9 IU/ml) with enzyme-linked immunosorbent assay (ELISA), as previously described [23]. ELISA kits were used for assay of serum levels of total adiponectin (Bühlman, Basel, Switzerland), leptin (Quantikine; R&D Systems, Oxford, UK), and visfatin/NAMPT (Bühlman, Basel, Switzerland). Insulinemia, used for calculation of the Homeostatic Model Assessment of Insulin Resistance (HOMA-IR) index, was assayed with chemiluminescence (ARCHITECT Insulin, Abbott Park, IL, USA).

Radiologic data

Radiographs of the hands and wrists (anteroposterior view) and the feet (anteroposterior and oblique views) were available at inclusion and at 1 year. The interpretation was standardized, as described previously [22,24]. Radiographs were read by the patient's office-based rheumatologist to determine the presence or absence of

abnormalities related to RA. Then, for quantitative scoring, radiographs were sent to the coordinating center for calculation of the total, joint erosion, and narrowing Sharp/van der Heijde score (SHS) at inclusion and 1 year by two experienced readers with blinding to patient data. Missing data were excluded from the analysis.

Statistical analysis

Continuous variables are described as mean \pm standard deviation (SD), and categorical variables as frequencies and percentages. The distributions of all variables were examined. Serum levels of adiponectin, leptin, visfatin/NAMPT, and CRP, Health Assessment Questionnaire (HAQ) score, and SHS values were not normally distributed and thus were log-transformed to remove positive skewness.

Baseline demographic, clinical, and radiographic characteristics of RA and UA patients were compared with χ^2 or Fisher Exact tests for discrete variables, and unpaired *t* test or Kruskal-Wallis test for continuous variables.

The association of adipokine levels and baseline radiographic damage was tested by univariate and multivariate linear regression analyses for patients with SHS >0 . Patients with SHS = 0 were removed from these analyses after logarithmic transformation.

Univariate and multivariate logistic regression analyses were used to model radiographic progression as a dichotomous variable, with the primary outcome defined by any change in total SHS (Δ SHS ≥ 1), as previously examined in the ESPOIR cohort [25,26]. Similarly, erosive and narrowing disease progression was defined as a Δ SHS ≥ 1 in erosive and narrowing subscores, respectively. Additional analyses involved a stricter outcome of any increase in total SHS ≥ 5 (Δ SHS ≥ 5), which delineates the subgroup of patients with rapid radiographic disease progression, also previously examined in the ESPOIR cohort [27].

Relevant known confounders were included in multivariate analyses models. We consider four models as follows: model 1, or “base” model, adjusted on age and sex; Model 2, or “metabolic” model, adjusted on Model 1 variables and body mass index (BMI) and HOMA-IR index; model 3, or “metabolic and RA” model, adjusted on model 2 variables and CRP level, DAS28, HAQ score, RF status, anti-CCP antibody status, and, for the association with radiographic progression, presence of typical damage related to RA at inclusion; and model 4, or “metabolic, RA, and steroid” model, adjusted on model 3 variables and steroid prescription at inclusion.

Sensitivity versus the false-positive frequency (1-specificity) for predicting radiographic disease progression (Δ SHS ≥ 1 or ≥ 5) with adipokines levels and with other

markers of radiographic disease progression (that is, DAS28 value, number of swollen joints, CRP level, RF level, and anti-CCP antibodies level) was analyzed with a receiver-operated characteristic (ROC) curve. The predictive accuracy of each item was assessed by using the area under the curve (AUC). To determine the optimal cut-off of adiponectin level, the Youden index was calculated by using the following formula: sensitivity + specificity - 1, and the maximum value of the Youden index corresponded to the optimal cut-off point [28].

All statistical tests were two-sided with a statistical significance defined as $P < 0.05$. SAS v9.3 (SAS Inst., Cary, NC, USA) was used for analysis.

Results

Characteristics of the study population and comparison of RA and UA patients

Among 813 patients enrolled in the whole cohort, 791 had data for serum adipokine levels and were analyzed in this study; 632 had RA fulfilling the ACR-EULAR 2010 criteria, and 159 had UA. Comparisons of clinical, biologic, and radiologic characteristics of RA and UA patients are shown in Table 1. At inclusion, RA patients were more often positive for anti-CCP antibodies and RF and showed radiographic changes than did UA patients. In addition, DAS28, HAQ score, and CRP level were significantly higher for RA than for UA patients. Baseline total SHS was higher for RA than for UA patients, the difference involving predominantly the erosive subscore. At 1-year follow-up, similar results were observed, with a wider variation of total and erosive SHS in the RA group.

Despite these differences, serum adipokine levels did not significantly differ between RA and UA patients (Table 1). However, adiponectin levels were higher for RA patients with positivity for anti-CCP antibodies or RF than for those without (5.22 ± 3.96 versus 4.58 ± 3.29 $\mu\text{g/ml}$; $P = 0.04$). Leptin level was higher but not significantly for patients without than for those with auto-antibodies (13.75 ± 12.70 versus 15.47 ± 15.35 ng/ml ; $P = 0.07$). Conversely, visfatin/NAMPT level did not vary by auto-antibody status ($P = 0.59$).

Serum adipokine levels and structural damage for patients with RA at baseline

Baseline serum adiponectin level was associated with baseline total SHS even after adjustment for metabolic and RA confounders, as well as steroid prescription (model 4: $\beta = 0.12$; $P = 0.006$) (Table 2). This association involved mainly the joint-narrowing subscore (model 4: $\beta = 0.14$; $P = 0.01$). Conversely, leptin and visfatin/NAMPT levels were not associated with total SHS on univariate or multivariate analysis (Table 2).

Table 1 Characteristics of patients with early rheumatoid arthritis (RA) and undifferentiated arthritis (UA) from the ESPOIR cohort

	UA (n = 159)	Early RA (n = 632)	P value
Age	47.2 ± 13.8	48.5 ± 12.2	0.46
Women (n,%)	117 (74%)	492 (78%)	0.25
First symptom (months)	6.6 ± 7.7	6.9 ± 8.5	0.72
DAS28	4.0 ± 1.0	5.4 ± 1.2	<0.0001
CRP level (mg/L)	17.2 ± 29.3	21.1 ± 33.1	0.0028
ESR	25.3 ± 22.4	30.6 ± 24.9	0.0014
Positive anti-CCP antibodies (n,%)	2 (1.26%)	313 (49.5%)	<0.0001
Positive RF (n,%)	5 (3.1%)	365 (57.75%)	<0.0001
Swollen-joint count	3.5 ± 2.4	8.2 ± 5.2	<0.0001
Tender-joint count	3.2 ± 2.6	9.9 ± 7.2	<0.0001
HAQ score	0.69 ± 0.58	1.05 ± 0.69	<0.0001
Diabetes mellitus (n,%)	7 (4.4%)	24 (3.8%)	0.73
HOMA-IR index	2.5 ± 2.6	2.8 ± 4.2	0.37
BMI	24.7 ± 4.6	25.2 ± 4.6	0.22
Steroid prescription at inclusion (n,%)	22 (13.8%)	84 (13.3%)	0.86
Radiographic damage at inclusion (n,%)	0 (0)	108 (17.1%)	<0.0001
Total SHS at inclusion	4.6 ± 6.8	6.14 ± 7.92	0.005
Erosive SHS	1.6 ± 3.13	3.09 ± 4.98	<0.0001
Narrowing SHS	2.97 ± 4.82	3.04 ± 4.36	0.77
Total SHS at 1 year	5.7 ± 9.5	7.7 ± 10.7	0.006
Erosive SHS	2.35 ± 5.05	4.48 ± 7.56	<0.0001
Narrowing SHS	3.36 ± 5.54	3.26 ± 4.78	0.9552
Total ΔSHS between inclusion and 1 year	1.0 ± 3.8	1.6 ± 4.4	0.02
Erosive ΔSHS between inclusion and 1 year	0.73 ± 2.7	1.37 ± 3.73	0.0025
Narrowing ΔSHS between inclusion and 1 year	0.24 ± 1.22	0.26 ± 1.15	0.7294
Adiponectin level (μg/ml)	4.9 ± 3.4	5.0 ± 3.7	0.63
Leptin level (ng/ml)	14.6 ± 14.4	14.4 ± 13.7	0.74
Visfatin/NAMPT level (ng/ml)	4.1 ± 3.6	3.57 ± 3.1	0.47

Data are expressed as mean ± SD unless indicated. Bold P values are significant.

DAS28, Disease Activity Score in 28 joints; CRP, C-reactive protein level; ESR, erythrocyte sedimentation rate; anti-CCP, anti-cyclic citrullinated protein peptide antibodies; RF, rheumatoid factor; HAQ, Health Assessment Questionnaire; VAS, visual analog scale; BMI, body mass index; HOMA-IR, Homeostatic Model Assessment of Insulin Resistance, SHS, total Sharp/van der Heijde score; ΔSHS, variation in SHS between inclusion and 1 year.

Association of adipokine levels and radiographic disease progression

Serum level of each adipokine are graphically reported in the Figure 1 according to the presence (n=160) or not of radiographic progression at 1 year (that is, ΔSHS ≥1 or ΔSHS <1).

Baseline serum adiponectin level was associated with radiographic disease progression (ΔSHS ≥1) in both univariate and multivariate models (Table 3, Figure 2). In Model 3 ("metabolic and RA"), baseline serum adiponectin level and radiographic progression at 1 year were related (adjusted odds ratio (OR), 95% confidence interval = 1.84 (1.24 to 2.71), P = 0.002). In other words, with each 1-unit increase in log(adiponectin) level, the risk of

radiographic progression at 1 year was increased 84%. This association persisted after adjustment for steroid prescription at inclusion (Model 4 in Table 3 and Figure 2).

Similarly, baseline serum adiponectin level was associated with erosive and narrowing disease progression (defined as ΔSHS erosive subscore ≥1 and ΔSHS narrowing subscore ≥1, respectively): adjusted (model 3) OR = 1.73 (1.17 to 2.55) (P = 0.006) and 1.91 (1.04 to 3.52) (P = 0.038), respectively (Table 3 and Additional file 1: Figure S1). This association persisted even after adjustment for steroid prescription at inclusion. No interaction was seen between adiponectin level and the two other adipokines levels, and the predictive value of adiponectin did

Table 2 Unadjusted and adjusted association of serum adipokine levels and total joint erosive and narrowing Sharp/van der Heijde score (SHS) at inclusion for RA patients with SHS ≥ 1 (n = 446)

		Univariate		Model 1 "Base"		Model 2 "metabolic"		Model 3 "metabolic + RA"		Model 4 "metabolic + RA + steroid"	
		β	P	β	P	β	P	β	P	β	p
Adiponectin	Total SHS	0.058	0.1	0.097	0.007	0.1	0.006	0.11	0.007	0.12	0.006
	Erosive SHS	-0.004	0.93	0.035	0.40	0.03	0.52	0.06	0.22	0.06	0.22
	Narrowing SHS	0.097	0.03	0.1	0.02	0.13	0.006	0.14	0.009	0.14	0.01
Leptin	Total SHS	0.015	0.75	0.015	0.73	-0.01	0.79	-0.01	0.76	-0.01	0.74
	Erosive SHS	-0.15	0.005	-0.13	0.01	-0.09	0.02	-0.06	0.14	-0.07	0.13
	Narrowing SHS	0.13	0.02	0.11	0.047	0.04	0.40	0.02	0.64	0.03	0.63
Visfatin/NAMPT	Total SHS	0.006	0.89	0.008	0.87	-0.01	0.82	-0.04	0.46	-0.04	0.46
	Erosive SHS	-0.08	0.11	-0.1	0.06	-0.12	0.03	-0.14	0.02	-0.14	0.02
	Narrowing SHS	-0.004	0.95	0.02	0.70	-0.009	0.89	-0.02	0.71	-0.02	0.71

Multivariate models: Model 1 ("base"): sex + age. Model 2 ("metabolic"): Model 1 + BMI + HOMA-IR index. Model 3 ("metabolic + RA"): Model 2 + CRP level + DAS28-ESR value + HAQ score + RF status + anti-CCP antibody status. Model 4 ("metabolic + RA + steroid"): Model 3 + steroid prescription at inclusion. The following variables were log-transformed: CRP level, HAQ score, serum adipokine levels. Results are presented as β regression coefficient and P value. Bold P values are significant.

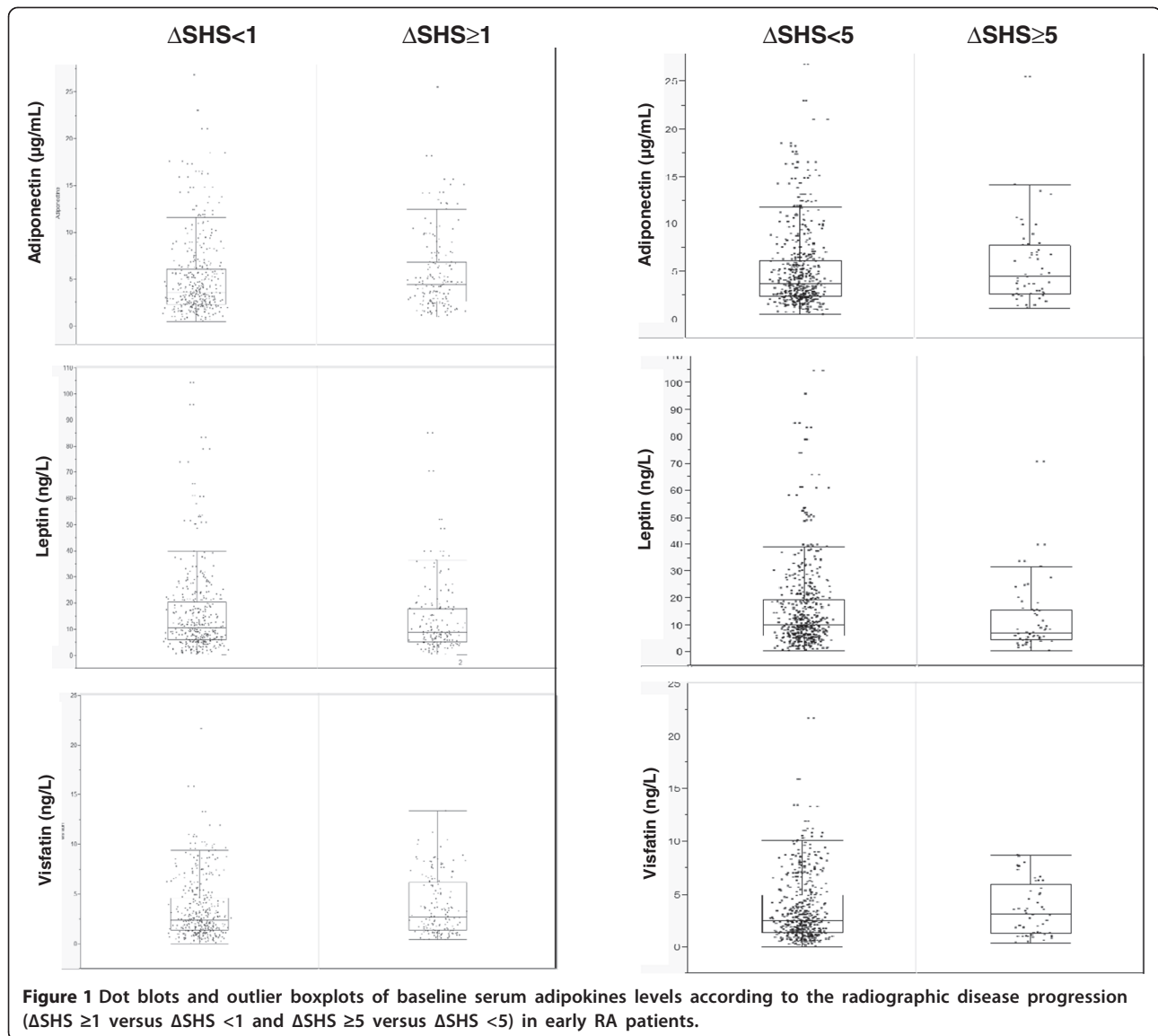


Figure 1 Dot blots and outlier boxplots of baseline serum adipokines levels according to the radiographic disease progression ($\Delta\text{SHS} \geq 1$ versus $\Delta\text{SHS} < 1$ and $\Delta\text{SHS} \geq 5$ versus $\Delta\text{SHS} < 5$) in early RA patients.

not change when serum leptin and visfatin/Nampt were included in the model 4 (data not shown).

We strengthened our results with a more-stringent definition of radiographic disease progression ($\Delta\text{SHS} \geq 5$; $n=54$). Serum levels of each adipokine according to the presence or absence of rapid radiographic progression are graphically reported in Figure 1. We still found an association of baseline serum adiponectin level and rapid radiographic progression even after adjustment for all considered confounders (model 4) (OR, 2.0 (1.14 to 3.52), $P = 0.02$) (Table 4, Figure 2).

For leptin level, with radiographic disease progression considered as $\Delta\text{SHS} \geq 1$ and after controlling on RA-related and metabolic parameters, with each 1-unit increase in $\log(\text{leptin})$ level, the risk of radiographic progression at 1 year was increased 59% (model 3 or 4, OR = 1.59 (1.05 to 2.41); $P = 0.03$) (Table 3, Figure 2). This association was

exclusively to the result of erosive disease progression (model 3 OR, 1.68 [1.1 to 2.56]; $P = 0.01$) (Table 3 and Additional file 1: Figure S1). These results did not change after controlling for steroid prescription (model 4). The association of leptin level and radiographic progression disappeared after adjustment when considering disease progression as $\Delta\text{SHS} \geq 5$ (Table 4, Figure 3).

To estimate the accuracy of serum leptin and serum adiponectin to independent prediction of radiographic disease progression at 1 year ($\Delta\text{SHS} \geq 1$ or $\Delta\text{SHS} \geq 5$), we performed ROC curves analyses of these two adipokines and also of other known markers associated with radiographic disease progression or rapid radiographic disease progression (/that is, DAS28 value, number of swollen joints, CRP level, RF level, and ACPA level) (Figure 4 for $\Delta\text{SHS} \geq 1$ and Figure 5 for $\Delta\text{SHS} \geq 5$). Interestingly, the AUC of each ROC curve of serum adiponectin and

Table 3 Association of baseline serum adipokine levels and radiographic disease progression defined as an increase of ≥ 1 in total SHS between inclusion and 1 year

		Univariate model		Model 1 "Base"		Model 2 "Metabolic"		Model 3 "Metabolic + RA"		Model 4 "Metabolic + RA + steroid"	
		OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	P
Adiponectin	Total Δ SHS ≥ 1	1.34 (1.03-1.76)	0.03	1.59 (1.19-2.12)	0.002	1.57 (1.16-2.13)	0.004	1.84 (1.24-2.71)	0.002	1.84 (1.25-2.72)	0.002
	Erosive Δ SHS ≥ 1	1.34 (1.02-1.75)	0.03	1.56 (1.17-2.08)	0.003	1.54 (1.13-2.08)	0.006	1.73 (1.17-2.55)	0.006	1.73 (1.17-2.55)	0.006
	Narrowing Δ SHS ≥ 1	1.65 (1.05-2.61)	0.03	1.79 (1.10-2.90)	0.02	1.95 (1.16-3.27)	0.01	1.91 (1.04-3.52)	0.04	1.93 (1.04-3.57)	0.04
Leptin	Total Δ SHS ≥ 1	0.86 (0.71-1.05)	0.14	0.95 (0.76-1.18)	0.63	1.24 (0.9-1.70)	0.2	1.59 (1.05-2.41)	0.03	1.59 (1.05-2.42)	0.03
	Erosive Δ SHS ≥ 1	0.88 (0.72-1.07)	0.19	0.96 (0.77-1.20)	0.69	1.29 (0.93-1.78)	0.12	1.68 (1.11-2.56)	0.01	1.69 (1.11-2.57)	0.01
	Narrowing Δ SHS ≥ 1	0.93 (0.66-1.3)	0.66	0.93 (0.64-1.35)	0.70	1.09 (0.63-1.87)	0.76	1.1 (0.58-2.09)	0.76	1.10 (0.58-2.08)	0.77
Visfatin/NAMPT	Total Δ SHS ≥ 1	1.22 (0.99-1.51)	0.06	1.20 (0.97-1.49)	0.09	1.2 (0.95-1.49)	0.12	1.09 (0.81-1.46)	0.57	1.09 (0.81-1.46)	0.58
	Erosive Δ SHS ≥ 1	1.25 (1.01-1.54)	0.04	1.23 (0.99-1.52)	0.06	1.22 (0.98-1.53)	0.08	1.14 (0.85-1.53)	0.37	1.14 (0.85-1.54)	0.37
	Narrowing Δ SHS ≥ 1	1.17 (0.82-1.67)	0.39	1.16 (0.81-1.67)	0.41	1.05 (0.73-1.53)	0.79	1.12 (0.72-1.75)	0.62	1.13 (0.72-1.77)	0.6

OR, odds ratio; 95% CI, 95% confidence interval. Total patient number, 530.

Data are unadjusted and adjusted odds ratios with p values. Bold P values are significant.

Radiographic progression is defined as variation in total SHS between inclusion and 1 year (total Δ SHS ≥ 1).

Erosive radiographic progression is defined as variation in erosive SHS between inclusion and 1 year (erosive Δ SHS ≥ 1).

Narrowing radiographic progression is defined as variation in narrowing SHS between inclusion and 1 year (narrowing Δ SHS ≥ 1).

Multivariate models:

Model 1 ("base"): sex + age (n = 530).

Model 2 ("metabolic"): Model 1 + BMI + HOMA-IR index (n = 497).

Model 3 ("metabolic + RA"): Model 2 + CRP level + DAS28-ESR value + HAQ score + RF status + anti-CCP antibody status + presence of radiographic changes at inclusion (n = 344).

Model 4 ("metabolic + RA + steroid"): Model 3 + steroid prescription at inclusion (n = 344).

The following variables were log-transformed: CRP level, HAQ score, serum adipokine levels.

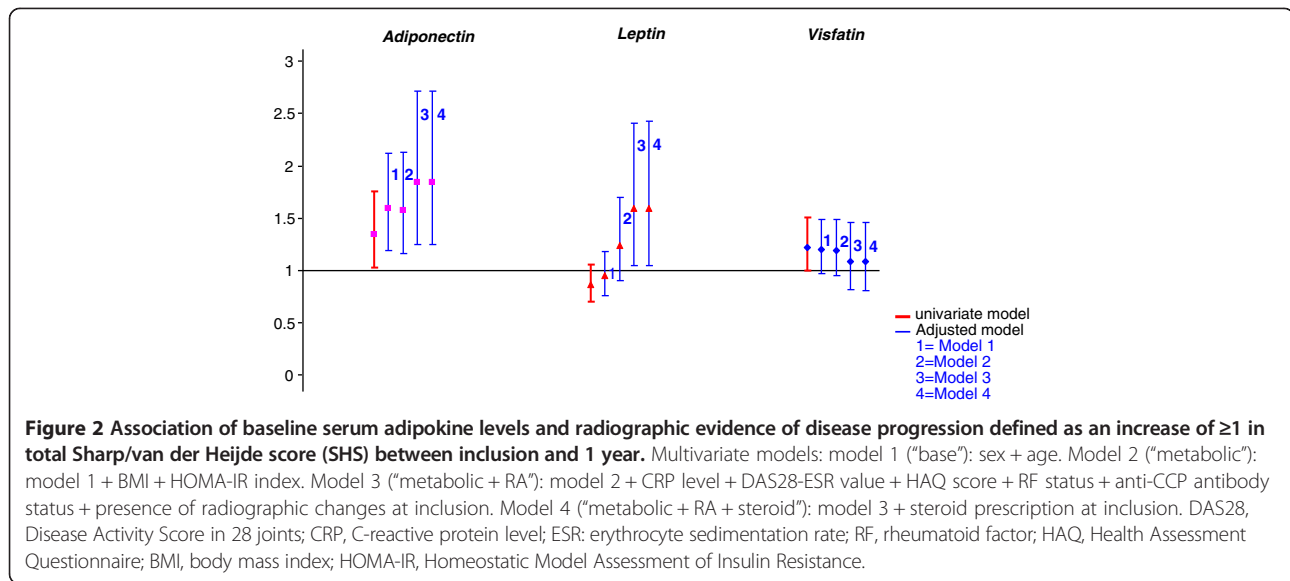


Figure 2 Association of baseline serum adipokine levels and radiographic evidence of disease progression defined as an increase of ≥ 1 in total Sharp/van der Heijde score (SHS) between inclusion and 1 year. Multivariate models: model 1 (“base”): sex + age. Model 2 (“metabolic”): model 1 + BMI + HOMA-IR index. Model 3 (“metabolic + RA”): model 2 + CRP level + DAS28-ESR value + HAQ score + RF status + anti-CCP antibody status + presence of radiographic changes at inclusion. Model 4 (“metabolic + RA + steroid”): model 3 + steroid prescription at inclusion. DAS28, Disease Activity Score in 28 joints; CRP, C-reactive protein level; ESR, erythrocyte sedimentation rate; RF, rheumatoid factor; HAQ, Health Assessment Questionnaire; BMI, body mass index; HOMA-IR, Homeostatic Model Assessment of Insulin Resistance.

serum leptin was in the same range as the AUC of baseline DAS28, number of swollen joints, or CRP level (P values nonsignificant). As expected and because of the accuracy of autoantibodies, RF level and anti-CCP antibodies level displayed higher AUC (P values < 0.05).

Considering the higher values of sensitivity combined with the lower values (1-specificity), the best cut-off for serum adiponectin level was 4.14 $\mu\text{g/ml}$ (sensitivity, 0.55, and specificity, 0.58) and 6.04 $\mu\text{g/ml}$ (sensitivity, 0.43, and specificity, 0.75) to predict $\Delta\text{SHS} \geq 1$ or ≥ 5 , respectively. We also calculated the negative predictive value (75% and 92% for $\Delta\text{SHS} \geq 1$ or $\Delta\text{SHS} \geq 5$, respectively) and the positive predictive value (36% and 16% for $\Delta\text{SHS} \geq 1$ or $\Delta\text{SHS} \geq 5$, respectively), suggesting that a nonelevation of serum adiponectin beyond the threshold may imply that the risk of radiographic disease progression is very low.

We used linear regression analyses to investigate the association of baseline serum adipokine levels and total SHS

at 1 year to ensure the association of serum adiponectin or leptin levels and radiographic disease progression. We focused on patients with severe disease, with prevalent radiographic lesions, because the log-transformed SHS automatically excluded all patients with SHS of 0. Again, serum adiponectin level predicted radiographic disease damage at 1 year (model 4, $\beta = 0.13$, $P = 0.0026$; Additional file 2: Table S1). Conversely, leptin level was not associated with total SHS at 1 year.

Baseline visfatin/NAMPT levels did not predict radiographic disease progression in any models, whatever the definition of radiographic disease progression used.

Discussion

We used data for the French ESPOIR cohort, one of the largest worldwide cohorts of early RA patients followed up longitudinally. We demonstrated that, in patients fulfilling the ACR-EULAR 2010 criteria for early RA at

Table 4 Association of baseline serum adipokine levels and rapid radiographic disease progression defined as an increase of ≥ 5 units in total SHS between inclusion and 1 year (total $\Delta\text{SHS} \geq 5$)

		Univariate model		Model 1 “Base”		Model 2 “metabolic”		Model 3 “metabolic + RA”		Model 4 “metabolic + RA + steroid”	
		OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	P
Adiponectin	$\Delta\text{SHS} \geq 5$	1.47 (0.98-2.20)	0.06	1.7 (1.11-2.61)	0.02	1.85 (1.16-2.94)	0.01	2.0 (1.14-3.52)	0.02	2.0 (1.14-3.52)	0.02
Leptin	$\Delta\text{SHS} \geq 5$	0.73 (0.54-0.97)	0.03	0.75 (0.55-1.04)	0.09	0.98 (0.61-1.56)	0.92	0.97 (0.55-1.68)	0.90	0.97 (0.56-1.68)	0.90
Visfatin/NAMPT	$\Delta\text{SHS} \geq 5$	1.14 (0.83-1.56)	0.41	1.12 (0.82-1.54)	0.48	1.11 (0.79-1.55)	0.56	0.96 (0.64-1.44)	0.85	0.96 (0.64-1.44)	0.85

OR, odds ratio; 95% CI, 95% confidence interval.

Multivariate models:

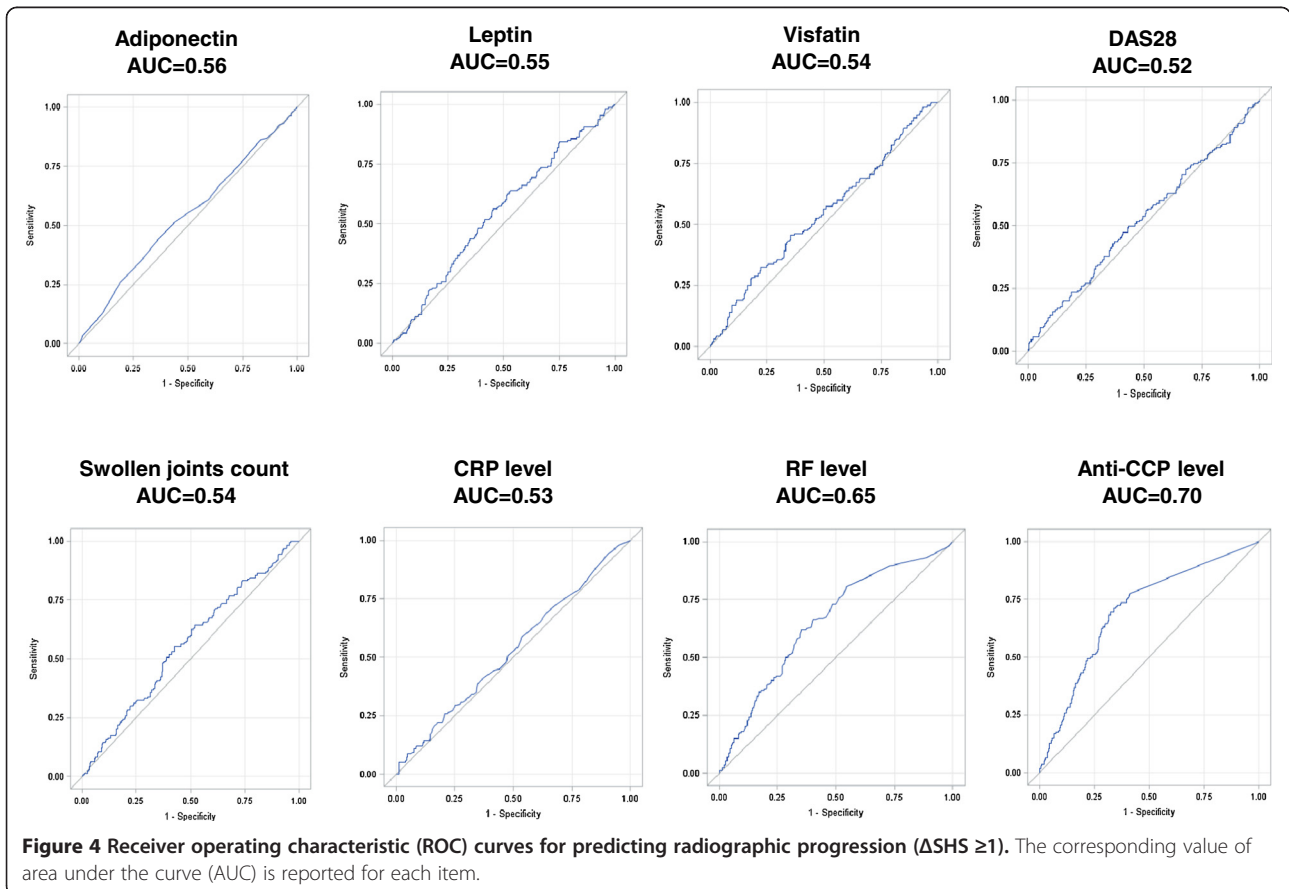
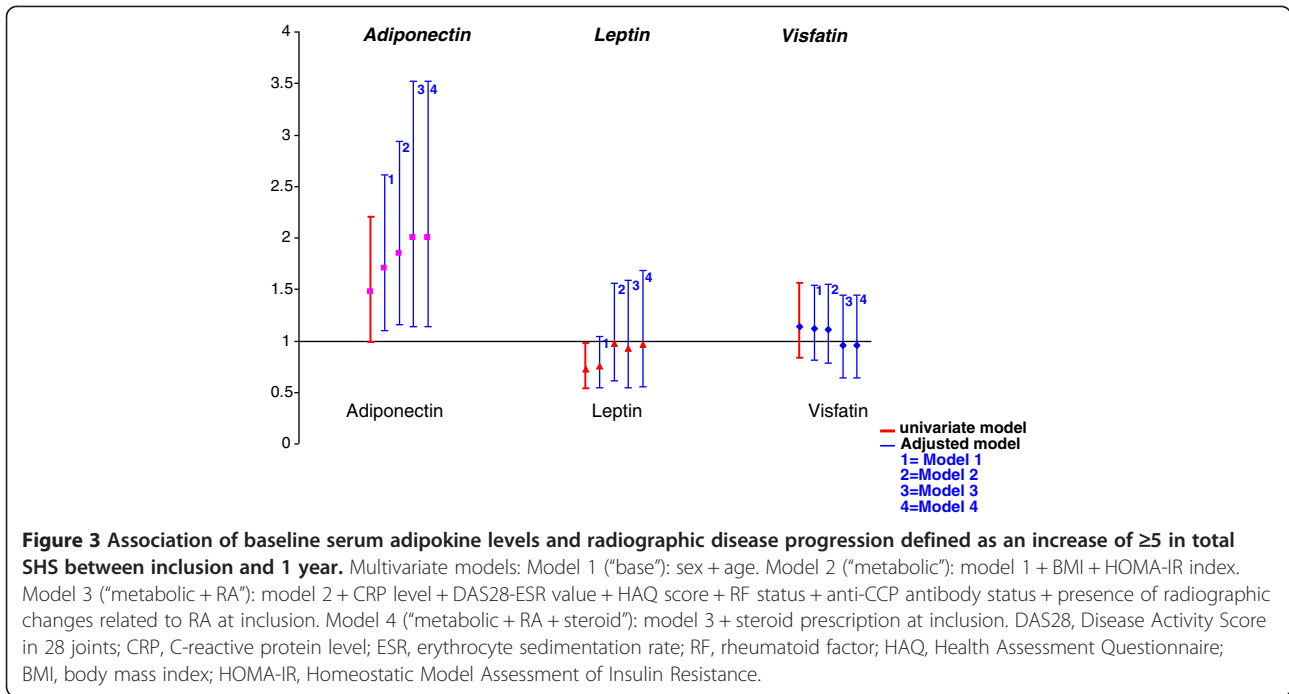
Model 1 (“base”): sex + age.

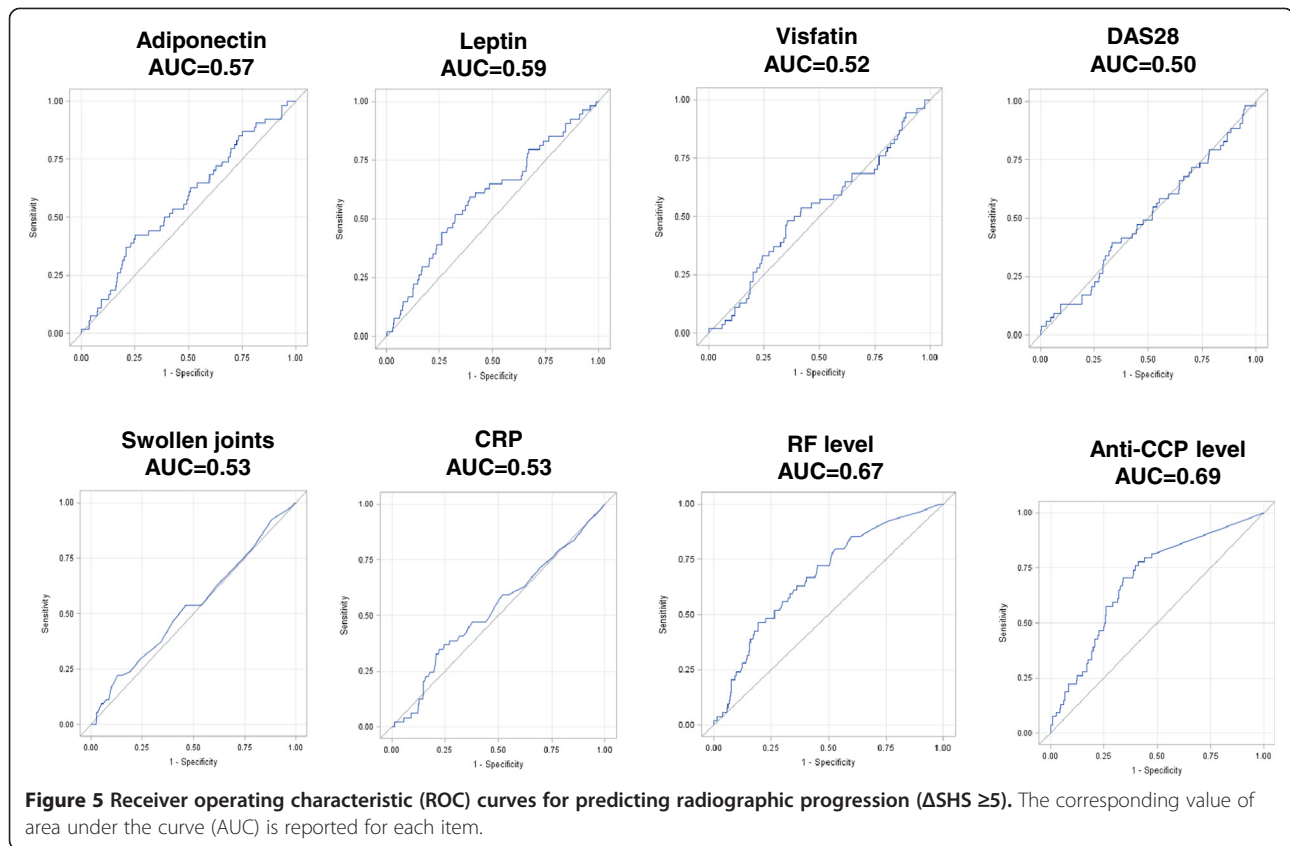
Model 2 (“metabolic”): model 1 + BMI + HOMA-IR index.

Model 3 (“metabolic + RA”): model 2 + CRP level + DAS28-ESR value + HAQ score + RF status + anti-CCP antibody status + presence of radiographic changes at inclusion.

Model 4 (“metabolic + RA + steroid”): model 3 + steroid prescription at inclusion.

The following variables were log-transformed: CRP level, HAQ score, serum adipokine levels. Bold P values are significant.





inclusion in the cohort, serum level of adiponectin is associated with subsequent early radiographic disease progression, independent of several RA and metabolic confounders and especially with rapid radiographic disease progression ($\Delta\text{SHS} \geq 5$). Serum adiponectin level is associated with both erosive and narrowing subscores of radiographic disease progression.

Klein-Wieringa *et al.* [1,20] investigated adipokine levels in 253 patients with early RA by using the 1987 ACR criteria, but we chose the new ACR-EULAR 2010 criteria, which do not involve radiographic results and thus are more accurate for early RA diagnosis and more adapted to identify new biomarkers of RA severity early during the disease course.

Several cross-sectional studies of established RA have shown a positive association of serum levels of adiponectin and visfatin/NAMPT and an inverse association of serum leptin level and radiographic joint damage [18,19]. However, we found that baseline serum adiponectin level was associated with quantitative joint damage, as assessed by SHS, mainly the joint-narrowing SHS subscore. Conversely, serum leptin was not associated with radiographic RA damage after multiple adjustments. Several reasons may explain such discrepancies. First, characteristics of the population differed because we investigated a large cohort of patients with early, not-established, untreated

RA, whereas Rho *et al.* [19] investigated 167 patients with established RA of 3-year duration. Second, we analyzed RA patients with $\text{SHS} \geq 1$, characterized by a more severe structural disease at inclusion, because of the logarithmic transformation of SHS necessary to meet the assumption of linear regression analysis. We found no association of leptin level and radiographic progression by total SHS; this finding is in agreement with that of Rho *et al.* [19], who showed that the association of serum leptin level and radiographic damage did not persist after adjustment for BMI, considered an important confounder.

Several of our statistical analyses show that adiponectin and leptin can predict radiographic progression and thus disease severity. Although adiponectin could have protective roles in some tissues [8], preclinical studies have demonstrated its deleterious role in cartilage homeostasis. Adiponectin induces the release of proinflammatory cytokines (that is, IL-6, monocyte chemoattractant protein-1 (MCP-1)), pro-degradative enzymes (that is, matrix metalloproteinases (MMPs)) and nitric oxide in chondrocytes [29-33]. These experimental data may explain, at least in part, a link between serum adiponectin level and joint-space narrowing found in our study. Moreover, synovial fibroblasts express adiponectin receptors and are reactive to adiponectin stimulation, triggering a proinflammatory and prodegradative state characterized by

prostaglandin E₂, IL-6, IL-8, MCP-1, and MMPs releases [29,33,34].

Furthermore, the paradoxical adiponectin effects between the cardiovascular system and joints may be related to its different isoforms. However, all adiponectin isoforms were shown to exert a proinflammatory and joint-destructive role on synovial cells [35]. Moreover, we previously compared levels of total and of the main isoform of adiponectin, "high-molecular-weight" (HMW) adiponectin in a large number of individuals with a wide range of BMI ($n = 408$). We found that total adiponectin and HMW-adiponectin were highly correlated ($r = 0.927$; $P < 0.001$, JP Bastard, personal communication). Therefore, total serum adiponectin level could be considered a good reflection of the HMW-adiponectin level. However, the direct assessment in the serum of the several adiponectin isoforms remains to be investigated in RA to compare its accuracy with serum total adiponectin level to predict structural progression.

Giles *et al.* [21] performed a cross-sectional study assessing the association of serum adipokine levels and radiographic damage and showed an association of high adiponectin level and erosions, as well as joint narrowing. Our study is the first to investigate the longitudinal association of serum adipokine levels and SHS subscores (that is, erosive and narrowing SHS). Here, we found that adiponectin level may predict joint erosive as well as narrowing disease progression at 1 year, which illustrates the long-term effect of this adipokine on all cell types within the RA joint [29-36].

We also found that the leptin level is a reliable surrogate biomarker of disease progression. The predictive value of leptin seems weaker than that of adiponectin, because we found no association of baseline leptin level and structural damage at inclusion of RA patients or subsequent rapid disease progression ($\Delta\text{SHS} \geq 5$). Interestingly, two longitudinal studies obtained similar results: no predictive value of serum leptin level for radiographic disease progression [20,21].

Considering the direct link between serum adipokine levels and metabolic disorders such as obesity, type 2 diabetes, insulin resistance, or metabolic syndrome [37-39], our interpretation of serum adipokine levels by multivariate analysis included not only BMI, but also an estimate of insulin resistance by the HOMA-IR index was not considered in previous studies [18-21]. Likewise, we included the HOMA-IR index among the "metabolic confounders" in multivariate models. Interestingly, the association of adiponectin or leptin level and radiographic disease progression persisted in these models. Adiponectin level may be independent of fat-tissue status in exerting a deleterious effect on RA joints, and thus is not the sole biologic explanation for the protective effect of obesity on RA joint damage [40,41].

Treatment at baseline, especially corticosteroids, may modify adipokine levels [42,43]. In the ESPOIR cohort, most patients were untreated at the time of inclusion, but about 14% received steroids for a short time before inclusion. We included steroid prescription as a confounder in our multivariate analysis (model 4). Steroid use modulates adipokine levels themselves [42,43]. Interestingly, steroid prescription did not modify the association of serum adipokine levels and radiographic changes, so use of steroids, given to patients with more severe disease, cannot explain the link between adipokine levels and RA progression.

We found no association of visfatin/NAMPT levels and RA, which disagrees with the cross-sectional study of Rho *et al.* [19] but agrees with data from a recent longitudinal study [20]. Thus, the use of this adipokine as serum biomarker seems more elusive than adiponectin.

One limitation of our study is that some concerns may be raised by the definition of radiographic progression as SHS ≥ 1 . However, to ensure the strength of our results, we performed two sensitivity analyses. First, we chose another more-stringent definition of radiographic progression ($\Delta\text{SHS} \geq 5$), which corresponds to the destruction of one small rheumatoid joint, thereby delineating rapid radiographic progression [27]. With this definition, never used in previous studies on serum adipokines [20,21], the associations persisted, which suggests that adiponectin may accurately differentiate slow and fast disease progression. Second, we performed a linear regression analysis searching for a correlation between baseline adipokine levels as continuous variables and SHS values at 1 year. Baseline adiponectin again was associated with structural damage at 1 year.

Since we identified serum adiponectin level as a predictive marker of subsequent structural disease progression, we have attempted to determine the best threshold by using ROC curves analyses (4.14 $\mu\text{g/ml}$ for $\Delta\text{SHS} \geq 1$ and 6.04 $\mu\text{g/ml}$ for $\Delta\text{SHS} \geq 5$). These two cut-offs display good specificity (58% and 75% for $\Delta\text{SHS} \geq 1$ and $\Delta\text{SHS} \geq 5$, respectively) and high negative predictive value (75% and 92% for $\Delta\text{SHS} \geq 1$ or $\Delta\text{SHS} \geq 5$, respectively), especially for rapid radiographic progression. Thus, a serum adiponectin below these two thresholds may exclude the risk of structural progression.

The next step for the validation of serum adiponectin is to model a matrix that would combine serum total adiponectin with other markers (anti-CCP antibodies status, CRP level, baseline swollen-joint count, and erosions seen on radiography) [27]. Furthermore, even if serum adiponectin measurement has drawbacks, such as low sensitivity and positive predictive value, its predictive accuracy is in the same range as other recognized predictive markers of radiographic progression, according AUC (that is, DAS28 value, number of swollen joints, and CRP level). Finally, the independent association between

serum adiponectin level and RA progression replicated here in such a large cohort gives pathophysiologic clues about the involvement of adipokines in the early clinical phase of RA and clearly demonstrates the link between a high level of serum adiponectin and structural disease progression.

Conclusions

We demonstrated that serum adiponectin level at the time of RA diagnosis represents a surrogate biomarker of early radiographic disease progression in patients fulfilling the new ACR-EULAR 2010 criteria and thus may accurately identify patients at high risk of early joint destruction independent of inflammation, autoantibody status, and metabolic disturbances.

Patient consent

Obtained.

Ethics approval

Approval was obtained from Montpellier University Ethics Committee.

Additional files

Additional file 1: Figure S1. Association of baseline serum adipokine levels and radiographic joint erosive or narrowing disease progression defined as an increase of ≥ 1 in erosive or narrowing SHS between inclusion and 1 year.

Additional file 2: Table S1. Association of serum adipokine levels and total Sharp/van der Heijde score (SHS) at 1 year.

Abbreviations

ACR: American College of Rheumatology; anti-CCP: Anti-cyclic citrullinated peptide; AUC: Area under the curve; BMI: Body mass index; CRP: C-reactive protein; ELISA: Enzyme-linked immunosorbent assay; ESR: Erythrocyte sedimentation rate; EULAR: European League Against Rheumatism; HAQ: Health Assessment Questionnaire; HOMA-IR: Homeostatic Model Assessment of Insulin Resistance; NAMPT: Nicotinamide phosphoribosyltransferase; OR: Odds ratio; RA: Rheumatoid arthritis; RF: Rheumatoid factor; ROC: Receiver-operated characteristic; SD: Standard deviation; SHS: Sharp/van der Heijde score; UA: Undifferentiated arthritis; Δ SHS: Change in total SHS.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

Conception and design: MM, JS, JC, TS, and FB. Acquisition of data: SF, J-PB, and JC. Analyses and interpretation of data: MM, JS, SF, SK, J-PB, OM, FL, TS, JC, and FB. Drafting the manuscript: MM, JS, SK, and FB. Revising the manuscript: MM, JS, SF, SK, J-PB, OM, FL, TS, JC, and FB. All authors read and approved the final manuscript.

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References

1. Aletaha D, Neogi T, Silman AJ, Funovits J, Felson DT, Bingham CO 3rd, Birnbaum NS, Burmester GR, Bykerk VP, Cohen MD, Combe B, Costenbader KH, Dougados M, Emery P, Ferraccioli G, Hazes JM, Hobbs K, Huizinga TW, Kavanaugh A, Kay J, Kvien TK, Laing T, Mease P, Ménard HA, Moreland LW, Naden RL, Pincus T, Smolen JS, Stanislawski-Biernat E, Symmons D, et al: **Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative.** *Arthritis Rheum* 2010, **2010**:2569–2581.
2. Lard LR, Visser H, Speyer I, vander Horst-Bruinsma IE, Zwinderman AH, Breedveld FC, Hazes JM: **Early versus delayed treatment in patients with recent-onset rheumatoid arthritis: comparison of two cohorts who received different treatment strategies.** *Am J Med* 2001, **111**:446–451.
3. Lindqvist E, Eberhardt K, Bendtzen K, Heinigard D, Saxne T: **Prognostic laboratory markers of joint damage in rheumatoid arthritis.** *Ann Rheum Dis* 2005, **64**:196–201.
4. van der Helm-van Mil AH, Verpoort KN, Breedveld FC, Toes RE, Huizinga TW: **Antibodies to citrullinated proteins and differences in clinical progression of rheumatoid arthritis.** *Arthritis Res Ther* 2005, **7**:R949–958.
5. Vittecoq O, Pouplin S, Krzanowska K, Jouen-Beades F, Menard JF, Gayet A, Daragon A, Tron F, Le Loet X: **Rheumatoid factor is the strongest predictor of radiological progression of rheumatoid arthritis in a three-year prospective study in community-recruited patients.** *Rheumatology (Oxford)* 2003, **42**:939–946.
6. Forslind K, Ahlmen M, Eberhardt K, Hafstrom I, Svensson B, Group BS: **Prediction of radiological outcome in early rheumatoid arthritis in clinical practice: role of antibodies to citrullinated peptides (anti-CCP).** *Ann Rheum Dis* 2004, **63**:1090–1095.
7. Lago F, Dieguez C, Gomez-Reino J, Gualillo O: **Adipokines as emerging mediators of immune response and inflammation.** *Nat Clin Pract Rheumatol* 2007, **3**:716–724.
8. Scotece M, Conde J, Gómez R, López V, Pino J, González A, Lago F, Gómez-Reino JJ, Gualillo O: **Role of adipokines in atherosclerosis: interferences with cardiovascular complications in rheumatic diseases.** *Mediators Inflamm* 2012, **2012**:125458.

9. Tilg H, Moschen AR: **Adipocytokines: mediators linking adipose tissue, inflammation and immunity.** *Nat Rev Immunol* 2006, **6**:772–783.
10. Heidemann C, Sun Q, van Dam RM, Meigs JB, Zhang C, Tworoger SS, Mantzoros CS, Hu FB: **Total and high-molecular-weight adiponectin and resistin in relation to the risk for type 2 diabetes in women.** *Ann Intern Med* 2008, **149**:307–316.
11. Bastard JP, Maachi M, Lagathu C, Kim MJ, Caron M, Vidal H, Capeau J, Fève B: **Recent advances in the relationship between obesity, inflammation, and insulin resistance.** *Eur Cytokine Netw* 2006, **17**:4–12.
12. Hascoet S, Elbaz M, Bongard V, Bouisset F, Verdier C, Vindis C, Annelise G, Taraszkiewicz D, Perret B, Galinier M, Carrière D, Ferrières J, Ruidavets JB: **Adiponectin and long-term mortality in coronary artery disease participants and controls.** *Arterioscler Thromb Vasc Biol* 2012, **33**:e19–29.
13. Jia T, Carrero JJ, Lindholm B, Stenvinkel P: **The complex role of adiponectin in chronic kidney disease.** *Biochimie* 2012, **94**:2150–2156.
14. Jorsal A, Tarnow L, Frystyk J, Lajer M, Flyvbjerg A, Parving HH, Vionnet N, Rossing P: **Serum adiponectin predicts all-cause mortality and end stage renal disease in patients with type I diabetes and diabetic nephropathy.** *Kidney Int* 2008, **74**:649–654.
15. Pham MN, Kolb H, Mandrup-Poulsen T, Battelino T, Ludvigsson J, Pozzilli P, Roden M, Schloot NC, Trial EC: **Serum adipokines as biomarkers of beta-cell function in patients with type 1 diabetes: positive association with leptin and resistin and negative association with adiponectin.** *Diabetes Metab Res Rev* 2013, **29**:166–170.
16. Lemoine M, Ratziu V, Kim M, Maachi M, Wendum D, Paye F, Bastard JP, Poupon R, Housset C, Capeau J, Serfaty L: **Serum adipokine levels predictive of liver injury in non-alcoholic fatty liver disease.** *Liver Int* 2009, **29**:1431–1438.
17. Neumann E, Frommer KW, Vasile M, Muller-Ladner U: **Adipocytokines as driving forces in rheumatoid arthritis and related inflammatory diseases?** *Arthritis Rheum* 2011, **63**:1159–1169.
18. Giles JT, Allison M, Bingham CO 3rd, Scott WM Jr, Bathon JM: **Adiponectin is a mediator of the inverse association of adiposity with radiographic damage in rheumatoid arthritis.** *Arthritis Rheum* 2009, **61**:1248–1256.
19. Rho YH, Solus J, Sokka T, Oeser A, Chung CP, Gebretsadik T, Shintani A, Pincus T, Stein CM: **Adipocytokines are associated with radiographic joint damage in rheumatoid arthritis.** *Arthritis Rheum* 2009, **60**:1906–1914.
20. Klein-Wieringa IR, van der Linden MP, Knevel R, Kwekkeboom JC, van Beelen E, Huizinga TW, van der Helm-van Mil A, Kloppenburg M, Toes RE, Ioan-Facsinay A: **Baseline serum adipokine levels predict radiographic progression in early rheumatoid arthritis.** *Arthritis Rheum* 2011, **63**:2567–2574.
21. Giles JT, van der Heijde DM, Bathon JM: **Association of circulating adiponectin levels with progression of radiographic joint destruction in rheumatoid arthritis.** *Ann Rheum Dis* 2011, **70**:1562–1568.
22. Combe B, Benessiano J, Berenbaum F, Cantagrel A, Daures JP, Dougados M, Fardellone P, Fautrel B, Flipo RM, Goupille P, Guillemin F, Le Loet X, Logeart I, Mariette X, Meyer O, Ravaud P, Rinceval N, Saraux A, Schaevebeke T, Sibilia J: **The ESPOIR cohort: a ten-year follow-up of early arthritis in France: methodology and baseline characteristics of the 813 included patients.** *Joint Bone Spine* 2007, **74**:440–445.
23. Gottenberg JE, Miceli-Richard C, Ducot B, Goupille P, Combe B, Mariette X: **Markers of B-lymphocyte activation are elevated in patients with early rheumatoid arthritis and correlated with disease activity in the ESPOIR cohort.** *Arthritis Res Ther* 2009, **11**:R114.
24. Devauchelle-Pensec V, Berthelot JM, Jousse S, Samjee I, Josseaume T, Colin D, Chales G, Thorel JB, Hoang S, Martin A, Youinou P, Le Goff P, Saraux A: **Performance of hand radiographs in predicting the diagnosis in patients with early arthritis.** *J Rheumatol* 2006, **33**:1511–1515.
25. Gottenberg JE, Dayer JM, Lukas C, Ducot B, Chiochia G, Cantagrel A, Saraux A, Roux-Lombard P, Mariette X: **Serum IL-6 and IL-21 are associated with markers of B cell activation and structural progression in early rheumatoid arthritis: results from the ESPOIR cohort.** *Ann Rheum Dis* 2012, **71**:1243–1248.
26. Sellam J, Kotti S, Fellahi S, Bastard JP, Meyer M, Lioté F, Meyer O, Simon T, Capeau J, Berenbaum F: **Serum hepcidin level is not an independent surrogate biomarker of disease activity or severity in early rheumatoid arthritis: results from the ESPOIR cohort.** *Ann Rheum Dis* 2013, **72**:312–314.
27. Fautrel B, Granger B, Combe B, Saraux A, Guillemin F, Le Loet X: **Matrix to predict rapid radiographic progression of early rheumatoid arthritis patients from the community treated with methotrexate or leflunomide: results from the ESPOIR cohort.** *Arthritis Res Ther* 2012, **14**:R249.
28. Youden WJ: **An index for rating diagnostic tests.** *Cancer* 1950, **3**:32–35.
29. Ehling A, Schaffler A, Herfarth H, Tarner IH, Anders S, Distler O, Paul G, Distler J, Gay S, Scholmerich J, Neumann E, Müller-Ladner U: **The potential of adiponectin in driving arthritis.** *J Immunol* 2006, **176**:4468–4478.
30. Frommer KW, Zimmermann B, Meier FM, Schroder D, Heil M, Schaffler A, Buchler C, Steinmeyer J, Brentano F, Gay S, Müller-Ladner U, Neumann E: **Adiponectin-mediated changes in effector cells involved in the pathophysiology of rheumatoid arthritis.** *Arthritis Rheum* 2010, **62**:2886–2899.
31. Tong KM, Chen CP, Huang KC, Shieh DC, Cheng HC, Tzeng CY, Chen KH, Chiu YC, Tang CH: **Adiponectin increases MMP-3 expression in human chondrocytes through AdipoR1 signaling pathway.** *J Cell Biochem* 2011, **112**:1431–1440.
32. Lago R, Gomez R, Otero M, Lago F, Gallego R, Dieguez C, Gomez-Reino JJ, Gualillo O: **A new player in cartilage homeostasis: adiponectin induces nitric oxide synthase type II and pro-inflammatory cytokines in chondrocytes.** *Osteoarthritis Cartilage* 2008, **16**:1101–1109.
33. Chen X, Lu J, Bao J, Guo J, Shi J, Wang Y: **Adiponectin: a biomarker for rheumatoid arthritis?** *Cytokine Growth Factor Rev* 2013, **24**:83–89.
34. Kusunoki N, Kitahara K, Kojima F, Tanaka N, Kaneko K, Endo H, Suguro T, Kawai S: **Adiponectin stimulates prostaglandin E(2) production in rheumatoid arthritis synovial fibroblasts.** *Arthritis Rheum* 2010, **62**:1641–1649.
35. Frommer KW, Schaffler A, Buchler C, Steinmeyer J, Rickert M, Rehart S, Brentano F, Gay S, Muller-Ladner U, Neumann E: **Adiponectin isoforms: a potential therapeutic target in rheumatoid arthritis?** *Ann Rheum Dis* 2012, **71**:1724–1732.
36. Luo XH, Guo LJ, Xie H, Yuan LQ, Wu XP, Zhou HD, Liao EY: **Adiponectin stimulates RANKL and inhibits OPG expression in human osteoblasts through the MAPK signaling pathway.** *J Bone Miner Res* 2006, **21**:1648–1656.
37. Arita Y, Kihara S, Ouchi N, Takahashi M, Maeda K, Miyagawa J, Hotta K, Shimomura I, Nakamura T, Miyaoka K, Kuriyama H, Nishida M, Yamashita S, Okubo K, Matsubara K, Muraguchi M, Ohmoto Y, Funahashi T, Matsuzawa Y: **Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity.** *Biochem Biophys Res Commun* 1999, **257**:79–83.
38. Okamoto Y, Kihara S, Funahashi T, Matsuzawa Y, Libby P: **Adiponectin: a key adipocytokine in metabolic syndrome.** *Clin Sci (Lond)* 2006, **110**:267–278.
39. Wannamethee SG, Lowe GD, Rumley A, Cherry L, Whincup PH, Sattar N: **Adipokines and risk of type 2 diabetes in older men.** *Diabetes Care* 2007, **30**:1200–1205.
40. van der Helm-van Mil AH, van der Kooij SM, Allaart CF, Toes RE, Huizinga TW: **A high body mass index has a protective effect on the amount of joint destruction in small joints in early rheumatoid arthritis.** *Ann Rheum Dis* 2008, **67**:769–774.
41. Westhoff G, Rau R, Zink A: **Radiographic joint damage in early rheumatoid arthritis is highly dependent on body mass index.** *Arthritis Rheum* 2007, **56**:3575–3582.
42. Fardet L, Antuna-Puente B, Vatier C, Cervera P, Touati A, Simon T, Capeau J, Fève B, Bastard JP: **Adipokine profile in glucocorticoid-treated patients: baseline plasma leptin level predicts occurrence of lipodystrophy.** *Clin Endocrinol (Oxf)* 2013, **78**:43–51.
43. Sukumaran S, Dubois DC, Jusko WJ, Almon RR: **Glucocorticoid effects on adiponectin expression.** *Vitam Horm* 2012, **90**:163–186.

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